THE CANADIAN ENTOMOLOGIST



Management of *Delia spp* (Diptera: Anthomyiidae) through selectively timed planting of *Phaseolus vulgaris* (Fabaceae) in Atlantic Canada.

Journal:	The Canadian Entomologist
Manuscript ID	TCE-ART-2018-009.R2
Manuscript Type:	Original Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Silver, Natalie; Government of Canada, Agriculture and Agri-Food Canada Hillier, Kirk; Acadia University, Biology Blatt, Suzanne; Agriculture and Agri-Food Canada, Entomology
Abstract:	Delia spp., including Delia platura (Meigen) (Diptera: Anthomyiidae) are known pests of vegetable crops. Here, three studies were conducted to examine the relationship between Delia spp. and Phaseolus vulgaris L. (Fabaceae). Field studies documented a relationship between planting data and occurrence of Delia spp. damage on P. vulgaris. Plantings in mid-June resulted in higher crop yields (mean bean pods per plant) and reduced damage ratings compared with earlier plantings. Late-May and early-June planting dates were not favorable, as they resulted in high damage ratings and low plant survival. Late-June and July plantings resulted in low damage ratings but low crops yields. Growth chamber experiments examined oviposition preference of D. platura females at seven phenological stages for two varieties of P. vulgaris. Results indicated significantly higher oviposition rates on bean plants at early phenological growth stages, with no significant varietal preference shown by maggots. Laboratory experiments quantified the impact of D. platura larval infestation on two vulgaris varieties at two growth stages. Results indicated no significant difference in variety choice. Recommendation for planting P. vulgaris to coincide with Delia spp. phenology using degree day model is discussed

SCHOLARONE*
Manuscripts

Cambridge University Press

 $\phi_{i \times j, \tau_{j}^{i}}$

Management of *Delia* spp. (Diptera: Anthomyiidae) through selectively timed planting of *Phaseolus vulgaris* (Fabaceae) in Atlantic Canada.

Natalie Silver¹, Kirk Hillier² and Suzanne Blatt^{3*}

¹ 24 Eden Row, Wolfville, Nova Scotia; 1 (902) 670-8434; natalie.m.silver@gmail.com

² Acadia University; 1 (902) 585-1314; kirk.hillier@acadiau.ca

³ Agriculture and Agri-Food Canada; 1 (902) 364-8552;

suzanne.blatt@arg.gc.ca

*corresponding author

1	Abstract	
2	Delia spp., including Delia platura (Meigen) (Diptera: Anthomyiidae)	
3	are known pests of vegetable crops. Here, three studies were	
4	conducted to examine the relationship between Delia spp. and	
5	Phaseolus vulgaris L. (Fabaceae). Field studies documented a	
6	relationship between planting date and occurrence of Delia spp.	
7	damage on P. vulgaris. Plantings in mid-June resulted in higher crop	
8	yields (mean bean pods per plant) and reduced damage ratings	
9	compared with earlier plantings. Late-May and early-June planting	
10	dates were not favorable, as they resulted in high damage ratings and	
11	low plant survival. Late-June and July plantings resulted in low	
12	damage ratings but low crops yields. Growth chamber experiments	
13	examined oviposition preference of D. platura females at seven	
14	phenological stages for two varieties of P. vulgaris. Results indicated	
15	significantly higher oviposition rates on bean plants at early	
16	phenological growth stages, with no significant varietal preference	
17	shown by maggots. Laboratory experiments quantified the impact of	
18	D. platura larval infestation on two P. vulgaris varieties at two	Comment [SB1]: Comment 1
19	growth stages. Results indicated no significant difference in variety	
20	choice. Recommendation for planting P. vulgaris to coincide with	
21	Delia spp. phenology using degree day model is discussed.	

22	introduction
23	Flies within the genus Delia (Diptera: Anthomyiidae) are known
24	agricultural pests in Canada, Europe and the United States (van
25	Schoonhoven and Voyest 1991). The following Delia spp. are
26	commonly found in Nova Scotia and PEI: D. florilega (Zetterstedt),
27	D. antiqua (Meigen), D. platura (Meigen), and D. radicum
28	(Linneaus). Delia spp. larvae cause significant losses to commercial
29	crops including Brassica napus L. by feeding on the root tissues,
30	negatively affecting plant survival (Finch 1989; Broatch et al. 2006;
31	Dosdall et al. 1994). Delia platura, the seedcorn maggot, causes crop
32	losses in many vegetables, and can cause significant damage in
33	common bean, Phaseolus vulgaris L. (Broatch et al. 2006; Finch
34	1989; Soroka and Dosdall 2011).
35	Nova Scotia and Prince Edward Island support three
36	generations of Delia spp. in a growing season (Howard et al. 1994).
37	The first generation will emerge from the soil in early spring, feeding
38	on germinating seeds and the roots of established plants (Finch 1989).
39	The first and second generations in each growing season are reported
40	to cause the majority of damage to crops (Howard et al. 1994). To
41	allow offspring easy access to food, adult females oviposit on soil
42	near plants and germinating seeds (Finch 1989; Howard et al. 1994).

Females use olfactory cues as their primary method in choosing an oviposition site (Gouinguené and Städler 2006). A single adult female can oviposit between 50 and 270 eggs (Bennett et al. 2011; Soroka and Dosdall 2011). Delia spp. larvae will eclose approximately two days after oviposition (Throne and Eckenrode 1986). The developmental time for larvae is approximately two week during which feeding occurs on vegetation. After the larval stage, Delia spp. will pupate, emerging as adults approximately two weeks later. The documented degree days between oviposition of one generation to the next is 495 DD (Funderburk et al. 1984).

Traditional control of *Delia* spp. involves the use of insecticides, and has been shown to reduce plant damage, however, few options are available to organic growers (Montecinos *et al.* 1986; Valenciano *et al.* 2004). One method used to manage *Delia* species in canola in Alberta Canada is the cultural practice of selectively timed planting (Dosdall et al. 1996). Selectively timed planting can cause asynchrony between the phenology of the plant and pest. An optimal planting date would avoid damage from *Delia* larvae, while also optimizing crop yield. *Delia* adults typically emerge from the soil in early spring and will lay eggs on germinating seed over a two-week period. Larvae hatch from eggs and feed on newly germinated

Silver, Page 5

seedlings before pupating. Such a method, if timed appropriately, could be useful to snap bean producers in PEI and NS. The snap bean industry in NS and PEI is small with each province having less than 20 acres in production. Snap beans are produced for local markets with each grower planting between 1 and 3 acres. Growers typically start direct seeding bean at the end of May or early June, with additional sequential plantings every 10-14 days to ensure a continuous supply of fresh beans for the local market. At harvest, the plant is pulled from the field and the bean pods removed and placed in a bin for transport to the market. The pods are graded and any pods with bacterial spot or insect chewing are left in the field. Growers sow at a density of 4 cm between bean seeds. This spacing creates large plants each producing 15-20 bean pods.

While most snap bean diseases attack the pods, *Delia* species will feed on the germinating seed and taproot tissues and, in severe cases, larvae will chew through the plant stem and tunnel up the stalk (Vea and Eckenrode 1976b). With no available controls, growers are left with little recourse but to accept loss. Selectively timed planting could allow establishment and maturation of the bean plant, well before attack by *Delia* species. Alternatively, a later planting date could avoid the first generation of *Delia* species altogether (Dosdall *et*

Comment [SB2]: Comment 3

al. 1996; Vea and Eckenrode 1976b). Selectively timed plantings are not a widely-used practice as further study is required to perfect this management strategy to synchronize planting with local *Delia* spp. populations (Finch 1989; Dosdall *et al.* 1996).

This research examines the cultural practice of selectively timed planting for management of *Delia* species in the Canadian Atlantic provinces of Nova Scotia and Prince Edward Island (PEI) in organically managed common bean, *P. vulgaris*. The major objectives of this study are to: (1) observe the relationship between planting date and occurrence of *Delia* species damage on *P. vulgaris*, (2) document host oviposition preference of *D. platura* females on two varieties and eight growth stages of *P. vulgaris*; and (3) to quantify the impact of *D. platura* larvae (at two levels of infestation) on two growth stages of *P. vulgaris*.

Materials and Methods

Monitoring Delia spp.

In each field, over both years, four blue 'dry touch' sticky traps (12 x 14 cm; Solida, Quebec, Canada) were attached to wooden stakes (1 x 4 cm and 1 m tall) with a bull clip 30 cm off the ground, as recommended by Finch (1989), and placed within the plot (see Fig.

Silver, Page 7

1). Two additional traps were located East and West of the plot, in the surrounding vegetation approximately 10 m away to capture the population prior to field preparation. Traps were collected and replaced each week. Only flies within the genus *Delia* were sexed and identified to species using a *Delia* taxonomic key (Savage *et al.* 2016). *Delia platura* and *Delia florilega* Zetterstedt males were counted while *D. platura* and *D. florilega* females were pooled as they are not reliably distinguishable morphologically. As the larvae are not distinguishable at the species level, any damage observed in the field studies could not be attributed to either of the *Delia* species present. As such, any reference to damage in the field studies will be attributed to *Delia* species. Degree days were calculated from 1 January each year, with a base of 3.9° Celsius (Funderburk *et al.* 1984).

Field Observations – 2015

Grower observation of plant loss early in the season in previous years and again in 2015 prompted closer observation in two Prince Edward Island (PEI) fields: Milton (46.306521° N, 63.277758° W) and Loyalist (46.309193° N, 64.358008° W). These fields were commercial organic agricultural fields surrounded by natural

vegetation or other crops, e.g. radish, soybean or winter wheat. No
insecticides were applied and weeding was done by hand. As per
usual practice, P. vulgaris (Gold Rush and Lewis varieties, yellow
and green, respectively) from Vesey's seed (York, PEI) were sown on
various planting dates: 16 May, 26 May and 5 June in Loyalist and on
16 June, 27 June and 7 July in Milton (Table 2). A preliminary
sample of damaged plants from the 16 and 26 May plantings in
Loyalist found Delia species larvae. Plant counts were conducted
when the plants reached the second trifoliate, stage 14, using the
BBCH-scale developed by the Biologische Bundesanstalt,
Bundessortenamt and Chemical industry (BBCH) working group
(Meier 2001, Table 1) and again at flowering, stage 65, by randomly
selecting $20-2$ m long sections and counting the number of plants
present. At these growth stages it was not possible to distinguish
variety and the sampling occurred over all rows of the same planting
date. As the fields had sequential plantings, the dates for these
samplings ranged from 12 June to 20 August 2015. At harvest, the
different varieties were easily distinguished. A sample of 30 plants
per planting date and variety (where applicable) were collected from
each field by walking down every second or third row (depending
upon number of rows in the planting) and selecting a plant every 20

Silver, Page 9

steps. Number of bean pods per plant was counted and damage from	n
Delia spp. to the root was recorded as present or absent.	

Field Trials - 2016

Based on the results from the 2015 field observations, replicated trials were conducted in three fields in Nova Scotia and PEI in 2016. Fields in PEI were located at Saint Ann (46.438631° N, 63.406906° W) and Wilmot Valley (46.384709° N, 63.701667° W) and the Nova Scotia site was located in Canning (45.170202° N, 64.437527° W). All field sites were commercial organic agricultural fields surrounded by natural vegetation or other crops, e.g. radish, soybean or winter wheat. No insecticides were applied to any site and weeding was done by hand.

In Canning and Saint Ann, blocks measuring 2.5 by 7.5 meters were established and contained six parallel rows (spaced 30 cm); where the two outer rows served as guard rows, and the 4 inner rows were used for treatments (Fig. 1). The guard rows were sown by hand with Gold Rush variety beans (2-3 cm deep and 5 cm apart) while the four inner rows were divided into four sections, each 1.5 m in length to create 16 plots. Additional guard plants, also Gold Rush variety and 6 per row were sown on either end of the treatment area. A 4 x 4 Latin square design (4 replicates of 4 planting dates) was sown by

hand with Lewis seed on the following dates: 17 May, 31 May, 14

June, 28 June, 2016 for Canning and 25 May, 8 June, 22 June and 6

July, 2016 for Saint Ann. Planting dates were consistent with

commercial grower planting timing in each area. At Wilmot Valley,
the treatment block was planted in amongst the regular grower

plantings and was 2 rows wide and 16 plots (each 1.5 m in length)
into the field and sown with 30 seed of Lewis. Guard plants, variety

Gold Rush, 6 in each row, were sown at each end of the treatment

block. Planting dates were systematically randomized within each
replicate and occurred on 1 June, 15 June, 29 June and 13 July 2016.

Plants were checked once or twice weekly and counted to assess survival. Any plant showing any sign of damage was sampled and examined to determine the cause. At harvest, bean plants were removed from the field and brought to the lab. Number of bean pods per plant and damage rating were recorded from all sites, fresh weight of the plant and root diameter was recorded from Wilmot Valley and Saint Ann. Root damage caused by *Delia* spp., was identified and categorized on a scale of 0-5 (Dosdall *et al.* 1994, Howard *et al.* 1994).

Silver, Page 11

Laboratory Studies

Laboratory studies were conducted in a walk-in growth chamber (constant temperature of 20 ± 1 °C, 60 ± 5 % RH, and a 16:8 light/dark photoperiod) located at the Kentville Research and Development Center, Kentville, NS. *Phaseolus vulgaris* var. Gold Rush and Lewis and *D. platura* colonies were reared and all experiments conducted under these conditions.

Insects

Colonies were maintained in Bug Dorms measuring 30 x 30 x 30 cm (BioQuip, California, USA). Water was provided in 25 mL solo cups, with the lid cut to hold a 5 cm long dental wick. *Delia platura* pupae were obtained from Francois Fournier (Collège Montmorency, Laval, Québec, Canada) and placed in the Bug Dorms to develop. Upon eclosion, adults could mate and oviposit freely. Dry food, composed of sucrose, milk powder, yeast, and organic soy flour in a ratio of 10:10:2:1, was provided in a petri dish (McClanahan and Miller 1958). Oviposition substrate consisted of a petri dish with damp paper towel and a slice of rutabaga. Egg collection and rutabaga replenishment occurred twice weekly. Eggs were transferred to 500 mL plastic containers filled with 200 mL of larval diet (Ishikawa *et al.* 1983). Up to 200 eggs were added, and covered in

damp vermiculite. Larvae hatched, developed and formed pupae in the larval food container. Pupae were then transferred to a petri dish with paper towel and damp vermiculite and placed in a Bug Dorm.

Emerging adults were separated by generation.

Phaseolus vulgaris seeds were individually planted in pots (7.5 cm diam. by 10 cm deep) and watered daily. Bean seed required for studies at the germination stage were grown on a petri dish with damp paper towel and covered with vermiculite. Plants were selected for experimentation when they reached one of eight different phenological stages: 03, 07, 10, 12, 13, 21, 65, and 75 (Table 1).

Female Oviposition Preference

Choice experiments occurred in Bug Dorms under the same conditions used for maintaining the colony. Ten *D. platura* mating pairs were placed within a Bug Dorm with two bean plants (one Lewis and one Gold Rush) for 48 hours, and provided with a water source and adult food. Bean plants of the same phenological stage and planting date, of similar shape and colour were selected for each trial. Ten replicates for each of 8 phenological stages were completed for a total of 80 trials. Plants and flies were used only once.

Following removal of adult flies, the soil, roots, and stem were

229	examined for eggs and larvae, as described by Dosdall et al. (1994).
230	Counts of eggs and larvae were recorded for each plant.
231	Larval impact trial
232	Phaseolus vulgaris plants (see plant growth conditions above
233	at two phenological stages, 12 and 13, were exposed to zero (control)
234	five or ten D. platura larvae until flies emerged as adults. The trial
235	began between 5 July and 19 July 2016 at asynchronous intervals; all
236	plants were then grown to maturity (stage 75, 56 days after planting)
237	when all plants were harvested. After harvest, each plant was
238	processed by recording the number of bean pods, weight of each
239	plant, and root damage (using the 0-5 scale) from July 28 to Sept 13,
240	2016.
241	Statistical Analysis
242	R Studio software was used to conduct all statistical analyses (R
243	Studio 2015).
244	Field observations – 2015
245	As the planting dates were not repeated across the two fields
246	(all 'early' plantings were in Loyalist and all 'later' plantings were in
247	Milton) only the three planting dates within each field could be
248	compared. Plant density (as mean number of bean plants per 2 m

length), and percentage of surviving plants was analysed following transformation using asin(x) and ANOVA methods for differences between planting date (within each field), and between plant stage surveyed within each field and planting date. Mean number of bean pods per plant were analysed for differences between planting date and variety (where both varieties were planted) using ANOVA methods at $\alpha=0.05$.

Field trials - 2016

Percentage of surviving plants was transformed using asin (x) and ANOVA methods to evaluate differences between planting date for each site. Mean damage rating, number of bean pods per plant, fresh plant weight, bean pod weight and root diameter were evaluated for differences between planting date using ANOVA within each site. Post-hoc means separation was determined using Tukey's Honestly Significantly Different (HSD) test at $\alpha = 0.05$. Degree day values were calculated using a threshold of 3.9°C (Broatch et al. 2006), starting accumulations from 1 January, to standardize the *Delia* species phenology across field sites in 2016 and cross-reference with plantings.

Laboratory trials

Silver, Page 15

Female oviposition preference was analysed using two-way ANOVA to determine if bean variety and plant stage influenced number of eggs oviposited. Oviposition Preference Index (OPI) was calculated using (X-Z)/(X+Z) where X is number of eggs oviposited on Lewis var. plants and Y is number of eggs oviposited on Gold Rush var. plants to evaluate preference of females for Lewis against Gold Rush varieties (Little et al. 2017). Larval impact on phenological stage and variety was analysed using ordinal logistical regression.

Results

Monitoring Delia spp.

Delia platura and D. florilega were the most abundant species capture in both years (Fig. 2A, 2B and 2C). Delia radicum and D. antiqua were also present but in very low numbers (e.g. less than 10 per collection). Three generations of Delia spp. were clearly present in all fields. More males than females were captured early in the season (Figs. 2B and 2C) while more females than males were captured later in the season (Fig. 2A).

Comment [SB3]: Comment 4

Field observations - 2015

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

Plant density was significantly affected by planting date in Milton and Loyalist fields on Prince Edward Island (Table 2). In both fields, the first planting had significantly fewer plants per 2 m transect than the last planting when assessed at growth stage 14 ($F_{2,47} = 91.3$, P < 0.0001 and $F_{2,27} = 14.8$, P < 0.0001, Loyalist and Milton fields, respectively) and at growth stage 65 ($F_{2,67} = 6.96$, P = 0.002 and $F_{2,57}$ = 5.75, P = 0.005, Loyalist and Milton fields, respectively). The 26 May planting at Loyalist experienced the greatest plant loss (plant death) and was replanted on 16 June. This planting had the lowest plant counts at growth stage 14 (0.85 plants) while the 7 July planting in Milton had the highest plant counts (21.5 plants) (Table 2). At growth stage 65, the 16 June planting in Milton had the fewest plants per transect (9.8 plants) while the replant of the 26 May planting in Loyalist (16 June) had the highest plant count at stage 65 (16.6 plants). Within each planting date, plant density was significantly different between assessments in Loyalist for the 5 June planting $(F_{1,48} = 5.63, P = 0.02)$ and in Milton for the 16 June $(F_{1,28} = 40.0, P =$ 0.04), and 7 July plantings ($F_{1,28} = 27.7$, P < 0.0001), in each case showing a decrease in plant density.

Mean number of bean pods at harvest was significantly
different by planting date within each field. In Loyalist there were
only two plantings with harvest data and both varieties. The 16 May
planting for both Lewis and Gold Rush varieties produced
significantly fewer beans per plant than the 16 June planting (replant
of the 26 May planting) ($F_{1,182} = 144.82$, $P < 0.0001$ and $F_{1,182} =$
15.33, $P < 0.0001$, planting date and variety, respectively, Table 2). In
the Milton field, there was no significant difference between the bean
varieties ($F_{1,136} = 2.05$, $P = 0.16$) for mean number of bean pods/plant,
but planting date was significantly different ($F_{2,136} = 41.5$, $P < 0.0001$,
Table 2). Greatest number of bean pods/plant were harvested from the
27 June planting with an average of 17.6 bean pods from the Gold
Rush and 15.2 bean pods from the Lewis varieties. The lowest
number of bean pods was harvested from the 7 July planting with an
average of 5.4 bean pods per plant. Estimates of the number of bean
pods/acre show the 16 June (replant of the 26 May planting) in
Loyalist and the 27 June planting in Milton to produce the greatest
yields. Yields were estimated to be 1870 and 1539 x 1000 bean
pods/acre for Gold Rush and Lewis varieties in Loyalist and 1577 and
1357 x 1000 bean pods/acre for Gold Rush and Lewis varieties in
Milton.

Examination of the plant roots post-harvest found high percentages of plants showing damage from *Delia* feeding in the Milton field with > 70% of the roots showing damage across all plantings and varieties (Table 2). Lowest percentages of plants with damaged roots were in the Loyalist field and the 16 May planting, 15.1 and 28.1%, Gold Rush and Lewis varieties, respectively.

Field trials - 2016

Survival of bean plants was significantly different across planting dates in Wilmot Valley ($F_{3,11} = 6.65$, P = 0.008) and Saint Ann ($F_{3,12} = 10.11$, P = 0.001) but not Canning ($F_{3,12} = 0.98$, P = 0.43) (Table 3). At all sites, plant survival was highest for later planting dates. Percentage of plants surviving until harvest was significantly lower for the first and second planting dates, in both Saint Ann and Wilmot Valley with no plants surviving the first planting (1 June) in Wilmot Valley (Table 3). Recovery of the seed from these plots found rotten, partially germinated seed containing larvae. Larvae were reared to adulthood and identified as D. platura and D. florilega. The 1 June planting in Wilmot Valley was replanted on 29 July 2016.

A significant decrease in mean damage rating across planting dates was found in Wilmot Valley ($F_{3,11} = 48.07$, P < 0.0001) and Saint Ann ($F_{3,12} = 25.23$, P < 0.0001) but not Canning ($F_{3,12} = 3.49$, P

Silver, Page 19

= 0.05). Across all sites, Wilmot Valley had the highest mean damage as the first planting date here, 1 June, experienced severe damage and failure to survive *Delia* attack. Examination of the ungerminated seed showed evidence of *Delia* spp. feeding. Later plantings at this site had significantly lower mean damage ratings. A similar pattern of damage early in growing season was observed at Saint Ann and Canning fields.

The number of bean pods per plant were significantly different across planting dates in Saint Ann ($F_{3,12} = 8.05$, P = 0.003) but not in Canning ($F_{3,12} = 0.92$, P = 0.46) or Wilmot Valley ($F_{3,11} = 3.27$, P = 0.06) (Table 3). A higher number of bean pods were harvested from plantings occurring in the middle of the growing season with number of bean pods per plant being lower at the beginning of the season (22.4 and 6.7 pods per plant in Saint Ann, first and last planting, respectively). Mean bean pod weight was statistically different across planting dates at Saint Ann ($F_{3,12} = 8.41$, P = 0.003) and Wilmot Valley ($F_{3,11} = 31.64$, P < 0.0001). Lowest bean pod weights were observed on the last planting date for Wilmot Valley and the second last planting date at Saint Ann, 1.94 g and 3.03 g, respectively (Table 3). Highest bean pod weights were from the 25 May planting for

Saint Ann (6.13 g) and the 15 June planting for Wilmot Valley (4.03 g).

Larger plants were harvested from plantings occurring earlier in the season for both sites on PEI. Plant mass and root diameter were higher from early plantings, and lower from later plantings across both PEI field locations (plant mass Wilmot Valley, $F_{1,\,4}$ = 44.24, P < 0.0001; Saint Ann, $F_{1,4}$ = 50.02, P < 0.0001; root diameter, Wilmot Valley, $F_{1,\,4}$ = 3.54, P < 0.0001; Saint Ann, $F_{1,4}$ = 3.06, P < 0.0001).

Laboratory Trials

· 376

Delia platura females showed no significant variety preference during oviposition ($F_{1,150} = 0.14$, P = 0.71, Fig. 3). Oviposition Preference Index values ranged from -0.13 to 0.004 across plant stages, suggesting that for any given plant stage the difference in number of eggs laid was at most 13% more on Gold Rush over Lewis. When varieties were pooled, a significant difference in oviposition rate between phenological stages was disclosed ($F_{7,71} = 5.41$, P < 0.001, Fig. 3). Significantly higher oviposition occurred at phenological stages 3 and 7, two to four days post germination, when compared with later stages (P < 0.05). There were no significant differences in larval damage to varieties of P.

Silver, Page 21

vulgaris tested ($F_{1,7} = 0.55$, P = 0.48, Table 4). A significantly higher damage rating was observed on the roots of plants infested with larvae over the controls (P < 0.05, Residual Deviance 186.3, Akaike Information Criteria (AIC) 204.3). Lower damage ratings were experienced by plants infested with larvae at phenological stage 12, first true leaf, than on stage 13, third true leaf. Although compelling, this result was not statistically significant. Further, the larvae did not complete development and were not recovered from the pots at the end of the trial. We cannot say for certain that the feeding pressure within each treatment (5 or 10 larvae) was consistent throughout the trial. Plant loss from other factors, pathogens (n = 2) and lack of germination (n = 1), also occurred in the laboratory.

Discussion

Organic bean producers in Nova Scotia and Prince Edward

Island can experience severe plant loss early in the growing season. It is recognized that *D. platura* prefer to oviposit on early growth stage *P. vulgaris* (Gouinguené and Städler 2006), and *P. lunatus* (Weston and Miller (1989) and that the most vulnerable time for a bean plant is from planting to 3 – 4 weeks of growth (van Schoonhoven & Voyest 1991; Vea & Eckenrode 1976b). Lack of lignification during early development leaves *P. vulgaris* vulnerable to attack, while healthy,

Comment [BS4]: Comment 7

Comment [BS5]: Comment 8

lignified P. vulgaris experience little damage from Delia spp. (Turnock et al. 1992). In this study, examination of damaged P. vulgaris plants disclosed damage characteristic of Delia spp. feeding (Howard et al. 1994; Turnock et al 1992; Vea et al 1975), described as superficial chewing of the taproot tissues and chewing through the plant stem and tunneling up the stalk (Vea and Eckenrode 1976b and Finch 1989). Additionally, these fields had high Delia species trap catches during May and June (average 50 to 350 per trap per collection date), and Delia larvae were recovered from damaged roots and stems, making *Delia* spp. the likely cause for the crop losses observed by growers. Further, significant damage by D. platura to P. vulgaris was documented under controlled conditions with plants having damage similar to that observed in field studies. The Canning site had a heavier soil type and retained moisture more than the field sites on PEI. As a result, some seed failed to germinate and upon recovery were found rotting and infected with fungal pathogens. This type of damage accounted for approximately 2% of the plants in this trial and was easily distinguishable from damage by Delia species. Producers wanting to ensure a continuous supply of fresh beans for the market will often use sequential plantings, starting when

the ground has thawed and repeat every 10-14 days. Such plantings,

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

430

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

Silver, Page 23

typically done without consideration of Delia spp. phenology, are at risk of coinciding with peak larval eclosion from the egg. The importance of selecting a planting date to maximize crop growth while avoiding pest damage has been noted by others (Balasubramanian et al. 2004, van Schoonhoven and Voyest 1991, Valenciano et al. 2004). An optimal planting date would result in a high number of plants producing a large number of bean pods per plant. Many studies evaluating the use of selectively timed plantings will refer to a period of time based on the Julian calendar, e.g. mid-June or early summer (Dosdall et al. 1995). Selection of a Julianbased planting date faces challenges when transferred to a different region due to climatic issues, requiring the study to be repeated in each region where the crop-pest complex is present. We propose using the degree-day model for D. platura, the predominant Delia species present in these fields to best determine the planting date most likely to result in avoidance of the larvae.

Degree day models for *D. platura* have been developed in the United States (Funderburk *et al.* 1984) and Korea (Lee *et al.* 2000) using field observations. In the laboratory, developmental rates for *D. platura* and *D. florilega* were determined by Sanborn *et al.* (1982) and Throne and Eckenrode (1986). Such studies can produce a range of

base development temperatures, e.g. 0.6-7.9°C, creating the need to evaluate each model in a specific regions. The field study by Funderburk *et al.* (1984) and the laboratory study by Sanborn *et al.* (1982) both found the same development temperature for *D. platura*, e.g. 3.9°C. In Canada, Broatch *et al.* (2006) used this threshold and found it effective to predict *D. platura* populations in canola in western Canada. Previous work with *Delia* species in NS have found this threshold to reasonably predict *Delia* phenology.

The phenology of the female is more critical than the male when trying to predict and avoid damage from the larvae. For the Delia species in NS and PEI, comparison of D. platura and D. florilega female phenology and planting dates from 2016 within each site (Figure 4) shows how the various plantings were likely to be impacted by Delia larvae. The first planting in Wilmot Valley (Figure 4B) was decimated (0% survival) by Delia larvae. This planting occurred at 277 DD_{3.9} and at the peak flight of the first generation. That creates the situation where females have been emerging and ovipositing eggs on the soil for a few days, many have likely have started to eclose and larvae are ready to feed on any newly germinating seed. The second planting at Wilmot Valley (at 400 DD_{3.9}) experienced high plant death (~73%) as well, occurring when

Comment [BS6]: Comment 10

Comment [BS7]: Comment 11

474

475

476

477

478

479

480

481

482

483

484

485

486

487

488

489

490

491

492

493

Silver, Page 25

oviposition would be starting to decline, but the eggs and larvae are still eclosing and feeding. By the third planting date 665 DD_{3.9}), ** Delia larvae are starting to pupate and only 28% loss was realised. The fourth planting (765 DD_{3.9}) experienced 34% loss, occurring shortly after the second emergence peak, when larvae would again be seeking food. The fifth planting (1024 DD_{3.9}) experienced 19% loss. Similar patterns were found in Canning and Saint Ann (Figure 4A and 4C). When seeding occurred during the first generation emergence, newly germinating seed were at risk of attack. Seed planted later in the season, even if Delia populations were on the rise, had the advantage of warmer temperatures to advance the lignification process. Based on these results, it would be recommended that growers monitor the degree days and plant their bean seed to avoid the first and second generations, i.e. between 500 and 600 DD_{3.9} for the first planting in areas where populations are high or about 400 DD_{3.9} where populations are low (see Saint Ann, PEI, Figure 4C). Avoidance of the second generation would require planting around 900 DD_{3.9}. However, planting at this time means that plants will have shorter days and fewer heat units for development, as P. vulgaris has a relatively high temperature threshold for growth, e.g. 10°C (de Medeiros et al. 2000). Planting earlier than 300 DD_{3.9} results in the

Comment [BS8]: Comment 13

bean seed not having enough heat units to grow and produce lignin to withstand larval feeding.

494

495

496

497

498

499

500

501

502

503

504

505

506

507

508

509

510

511

512

513

514

Data collected from our field and laboratory experiments suggest a potential for P. vulgaris to tolerate Delia spp. attack. Tolerance, the ability of a plant to compensate quickly after attack or to withstand attack, has been observed in many crops and a variety of insect pests, including the common bean (Blatt et al. 2008; Ojwang 2010; Vea and Eckenrode 1976a). In our study, the Wilmot Valley field site showed no significant change in number of bean pods per plant between the 15 June and 29 July plantings even though damage rating was 3.4 for the 15 June planting where only 21% of the plants survived, compared with the 29 July planting with a mean damage rating of 1.6 and 71% of plants surviving. Laboratory experiments found severe D. platura damage on plants infested at early phenological stages of growth, while low damage ratings were realised from older P. vulgaris plants. Consistent with Turnock et al. (1992), such tolerance could be due to lignification of these older plants.

Field evidence from 2015 suggested that *Delia* spp. may exhibit a varietal preference, an observation supported by Vea and Eckenrode (1976a). However, no significant difference in oviposition

Silver, Page 27

or extent of damage between bean varieties was observed in the current laboratory studies. Oviposition site is chosen by *Delia* spp. female flies through the detection of plant volatiles by olfactory senses (Gouinguené and Städler 2006; Hough-Goldstein 1985; Spencer *et al* 1995). The lack of oviposition site preference found in this study could speak to similarity in olfactory profiles between the two varieties of bean. As such, there is no benefit to using one of these varieties over the other to reduce *Delia* damage. One issue which may have skewed our results is the large number of flies used (ten males and ten females per replicate). This may have caused competition between females for an oviposition site, possibly altering preferences.

Conclusion

Laboratory experiments found no varietal preference by D. platura females for either Lewis or Gold rush varieties of P. vulgaris. In the lab, high oviposition rates and high damage from D. platura occurred on early phenological plant stages. In the field, selectively timed planting to avoid the first and second generations of Delia can be an effective strategy to avoid loss from Delia spp. feeding. However, to obtain high crop yields, a balance must be achieved between late planting to manage pest damage and early planting to

ensure high yield. Using the phenology of *Delia* spp. as an indicator of when to plant will provide growers with a selective planting date where young *P. vulgaris* plants are not subject to high *Delia* spp. larvae. Further study to compare plantings based on Julian day with degree day recommendations are required to verify these results.

Acknowledgements

The authors wish to thank Red Soil Organics, Vermuelen
Farms, Alexanders Fresh Vegetables as well as Jenn and Sandy's
Organic Farm for use of field sites for this work. Many thanks are
extended to the PEI Horticultural Association, Andrew Ryan, Shelley
Adams, Joanne Driscoll, and Caitlyn vanKeymeulen for their
technical support. We would also like to thank the anonymous
reviewers for their helpful comments. Funding was provided by
Agriculture and Agri-Food Canada (A-base: Project ID 811) and
Acadia University.

551	References
552	Balasubramanian, P., Vandenberg, A., and Hucl, P. 2004. Planting date and
553	suboptimal seedbed temperature effects on dry bean establishment,
554	phenology and yield. Canadian Journal of Plant Science, 84(1): 31-36.
555	BBCH 2001. Growth stages of mono-and dicotyledonous plants. (U. Meier,
556	Ed.) (2nd ed.). Berlin, Germany: Federal Biological Research Centre
557	for Agriculture and Forestry.
558	Blatt, S. E., Smallegange, R. C., Hess, L., Harvey, J. A., Dicke, M., and van
559	Loon, J. J. A. 2008. Tolerance of Brassica nigra to Pieris brassicae
560	herbivory. Botany, 86(6): 641–648.
561	Broatch, J., Dosdall, L., Clayton, G., Haker, K., and Yang, R. 2006. Using
562	degree-day and logistic models to predict emergence patterns and
563	seasonal flights of the cabbage maggot and seed corn maggot (Diptera:
564	Anthomyiidae) in canola. Environmental Entomology, 35(5): 1166-
565	1177.
566	Dosdall, L. M., Herbut, M. J., and Cowle, N. T. 1994. Susceptibilities of
567	species and cultivars of canola and mustard to infestation by boot
568	maggots (Delia spp.) (Diptera: Anthomyiidae). The Canadian
569	Entomologist, 126(2): 251–260.

570	Dosdall, L. M., Herbut, M. J., Cowle, N. T., and Micklich, T. M. 1996. The
571	effect of seeding date and plant density on infestations of root maggots,
572	Delia spp. (Diptera: Anthomyiidae), in canola. Canadian Journal of
573	Plant Science, 76(1): 169–177.
574	Finch, S. 1989. Ecological considerations in the management of Delia pest
575	species in vegetable crops. Annual Review of Entomology, 34(1): 117-
576	137.
577	Funderburk, J. E., Higley, L. G., and Pedigo, L. P. 1984. Seedcorn maggot
578	(Diptera: Anthromyiidae) phenology in central Iowa and examination
579	of a thermal-unit system to predict development under field conditions.
580	Environmental Entomology, 13(1) : 105-109.
581	Gouinguené, S. P., and Städler, E. 2006. Oviposition in Delia platura
582	(Diptera, Anthomyiidae): the role of volatile and contact cues of bean.
583	Journal of Chemical Ecology, 32(7): 1399–1413.
584	Hough-Goldstein, J. A. 1985. Oviposition site selection by seedcorn maggot
585	flies (Diptera: Anthomyiidae) unaffected by prior experience or larval
586	host. Environmental Entomology, 14(3): 289–292.

587	Howard, R., Garland, J., and Seaman, W. 1994. Diseases and Pests of
588	Vegetable Crops in Canada: an illustrated compendium. Ottawa,
589	Ontario: Entomological Society of Canada. 554 pp.
590	Ishikawa, Y., Mochizuki, A., Ikeshoji, T., and Matsumoto, Y. 1983. Mass-rearing
591	of the onion and seed-corn flies, Hylemya antiqua and Hylemya platura
592	(Diptera: Anthomyiidae), on an artificial diet with antibiotics. Appl. Ent. Zool
593	18(1): 62–69.
594	Little, C. M., Chapman, T. W., Moreau, D. L., and Hillier, N. K. 2017.
595	Susceptibility of selected boreal fruits and berries to the invasive pest
596	Drosophila suzukii (Diptera: Drosophilidae). Pest Management
597	Science, 73(1): 160–166.
598	McClanahan, R. J., and Miller, L. A. 1958. Laboratory rearing of the seed-
599	corn maggot, Hylemya cilicrura (Rond.) (Diptera: Anthomyiidae). The
600	Canadian Entomologist, 90(6): 372–374.
601	de Medeiros, G.A., Arruda, F. B., Sakai, E., Fujiwara, M. and Boni, N.R.
602	2000. Vegetative growth and bean crop coefficient as related to
603	accumulated growing degree-days. Pesquisa Agropecuaria Brasileira,
604	35 (9) : 1733-1742.

605	Montecinos, M. T., Patricio Arretz, V., and Araya, J. E. 1986. Chemical
606	control of Delia platura in Phaseolus vulgaris with seed and soil
607	treatments in Chile. Crop Protection, 5(6): 427-429.
608	Ojwang, P. 2010. Genetic Studies on Host-Plant Resistance to Bean Fly
609	(Ophiomyia spp.) and Seed Yield in Common Bean (Phaseolus
610	vulgaris) under Semi-Arid Conditions. University of KwaZulu-Natal,
611	Pietermaritzburg, Republic of South Africa.
612	R Studio Team 2015. R Studio: Integrated Development for R. R Studio,
613	Inc., Boston, MA Available from http://www.rstudio.com/. [accessed
614	13 December 2016].
615	Savage, J., Fortier, A., Fournier, F., and Bellavance, V. 2016. Identification
616	of Delia pest species (Diptera: Anthomyiidae) in cultivated crucifers
617	and other vegetable crops in Canada. Canadian Journal of Arthropod
618	Identification, 29: 1-40.
619	Soroka, J., and Dosdall, L. 2011. Coping with root maggots in prairie canol-
620	crops. Prairie Soils and Crops, 4: 24–31.
621	Spencer, J. L., Candolfi, M. P., Keller, J. E., and Miller, J. R. 1995. Onion
622	fly, Delia antiqua, oviposition and mating as influenced by insect age

623	and dosage of male reproductive tract extract (Diptera: Anthomyiidae).
624	Journal of Insect Behavior, 8(5): 617–635.
625	Turnock, W. J., Timlick, B., Galka, B. E., and Palaniswamy, P. 1992. Root
626	maggot damage to canola and the distribution of $Delia$ spp. (Diptera:
627	Anthomyiidae) in Manitoba. The Canadian Entomologist, 124(1): 49-
628	58.
629	Valenciano, J. B., Casquero, P. A., and Boto, J. A. 2004. Evaluation of the
630	occurrence of bean plants (Phaseolus vulgaris L.) affected by bean seed
631	fly, Delia platura (Meigen), grown under different sowing techniques
632	and with different forms of pesticide application. Field Crops Research,
633	85(2): 103–109.
634	van Schoonhoven, A., and Voyest, O. 1991. Common Beans: Research for
635	Crop Improvement. Melksham, Wiltshire: Redwood Press.
636	Vea, E. V., and Eckenrode, C. J. 1976a. Resistance to seedcorn maggot in
637	snap bean. Environmental Entomology, 5(4): 735–737.
638	Vea, E. V., and Eckenrode, C. J. 1976b. Seed maggot injury on surviving
639	bean seedlings influences yield. Journal of Economic Entomology,
640	69(4): 545–547.

641	Vea, E. V., Eckenrode, C. J., and Webb, D. R. 1975. Seedcorn maggot
642	injury. Geneva, N.Y.: New York State Agricultural Experiment Station.
643	Weston, P. A., and Miller, J. R. 1989. Ovipositional responses of seedcorn
644	maggot, Delia platura (Diptera: Anthomyiidae), to developmental
645	stages of lima bean. Annals of the Entomological Society of America,
646	82(3): 387–392.
647	

Table 1. Selected phenological stages of *Phaseolus vulgaris* (BBCH, 2001).

Code	Description
0	Dry seed
3	Radicle emergence from seed coat (two days post germination)
7	Hypocotyl break through seed coat (four days post germination)
10	Cotyledons emerged, unfolded
12	First true leaf emerged, unfolded
13	First trifoliate leaf emerged, unfolded (third true leaf)
14	Second trifolate leaf emerged, unfolded (fourth true leaf)
21	Growth of first side shoot
65	Blossoms in Flower (over 50% of flowers open)
75	Bean harvest (over 50% of pods at full length)

649

650

1 **Table 2:** Mean plant density (± SE) at two growth stages, mean number (± SE) of bean pods per plant at harvest, estimated

yield/acre and percentage of plants at harvest showing damage from Delia species for Phaseolus vulgaris var. Gold Rush and

3 Lewis, sown at various dates at two locations on Prince Edward Island in 2015. Means within column (for plant density) and

means within column (Number bean pods/plant) and variety (Loyalist field only) with same letter not significantly different 4

5 using ANOVA methods followed by Tukeys HSD mean separation test (P < 0.05).

Comment [BS9]: Comment 15

			Plant density*	ensity*		Mean # bean	Estimated.	% plants
Field	Plant date	Harvest date	GS 14	GS 65	Variety	pods/plant (± SE)	pods/acre (x 1000)	damaged roots
Loyalist	16 May	12 Aug	Loyalist 16 May 12 Aug 10.6 (0.97) a 11.5 (0.62) a	11.5 (0.62) a	Gold Rush	6.8 (0.79) a	524	15.2
					Lewis	3.3 (0.45) A	247	28.1
	26 May	24 Sep	0.8(0.32)b	16.6 (0.99) b	Gold Rush	16.9 (0.91) b	1870	44.1
					Lewis	13.9 (0.72) B	1539	55.7
	5 June	‡	18.1 (1.33) c	18.1 (1.33) c 15.1 (0.53) b	Gold Rush			45.0
Milton	16 June	4 Sep	11.3 (0.93) A	9.8 (0.65) A	Gold Rush	9.9 (1.60) A	584	78.9
	27 June	4 Sep	14.4 (1.86) A	14.4 (1.86) A 13.5 (1.47) B	Gold Rush	17.6 (1.62) B	1577	73.9
					Lewis	15.2 (1.20) B	1357	86.8
	7 July	12 Sep	21.5 (1.09) B	21.5 (1.09) B 13.3 (0.96) B	Lewis	5.4 (0.65) C	473	83.3

Plant densities taken from fields when plants too young to allow variety differentiation

¹Estimated yield determined by multiplying the mean number of bean pods/plant by the plant density at growth stage 65 and using a conversion factor of 6634 which is based upon a row spacing of 0.61 m and assuming that the plant density is the same for both

varieties

Cambridge University Press

*May 26 planting was replanted on June 16, plant density from stage 14 is the May 26 planting and from stage 65 is the June16 replacement planting **Grower harvested bean pods on 10 September before sample could be taken, leaving plants in the field. A sample of these plants allowed evaluation of *Delia* species damage for this planting 11 12 13

Silver, Page 38

Table 3. Mean (± SE) percent survival, mean (± SE) number of bean pods per plant, and mean (± SE) damage rating of

14

Phaseolus vulgaris at all field locations in NS and PEI in 2016. Letters within column, for each site, denote significant 15

differences within each measured variable using ANOVA methods and Tukeys HSD mean separation test (P < 0.05). 16

Comment [BS10]: Comment 15

	Tanting	reacent prant	Damage	Number Dean	LIMBIL	bean pod	Koot diameter
Field	date	survival	rating	pods/plant	mass (g)	weight (g)	(mm)
Canning,	17 May	55.0 (9.5)	2.4 (0.3)	4.9 (0.5)	:	1	1
SN	31 May	55.0 (9.5)	1.5 (0.3)	7.4 (1.9)	1	i	1
	14 June	70.0 (23.8)	1.9 (1.0)	5.9 (2.2)	:	ł	1
	28 June	75.0 (5.0)	0.0 (0.0)	4.1 (0.1)		ŀ	1
Wilmot	1 June*	0.0 (0.0)	ł	:		ı	ŀ
Valley,	15 June	20.8 (5.2) B	3.4 (0.2) A	18.9 (2.0)	189.31 (2.9) A	4.28 (0.8) A	8.50 (0.7) A
PEI	29 June	71.8 (4.2) A	1.6 (0.1) B	20.2 (0.8)	101.67 (12.0) C	1.78 (0.2) C	6.76 (0.1) BC
	13 July	65.6 (6.4) AB	1.3 (0.1) BC	23.4 (0.8)	137.43 (8.3) B	3.11 (0.3) B	7.00 (0.1) AB
	29 July	80.5 (9.2) A	0.5 (0.1) C	18.4 (0.7)	72.54 (7.4) C	1.94 (0.2) C	5.84 (0.1) BC
Saint	25 May	16.7 (4.1) b	3.4 (0.4) a	22.4 (5.1) a	237.32 (69.6) a	6.13 (0.5) a	9.45 (0.8) a
Ann,	8 June	20.0 (4.9) b	3.7 (0.3) a	21.6 (1.4) a	188.65 (23.0) ab	5.01 (0.6) a	8.40 (0.4) a
PEI	22 June	64.2 (11.4) a	1.4(0.2) b	9.2 (2.1) b	54.29 (17.0) b	3.03 (0.2) b	5.43 (0.5) b
	6 July	70.0 (4.3) a	0.6 (0.2) b	6.7 (0.7) b	57.31 (8.3) b	4.93 (0.3) a	5.38 (0.3) b

18

17

Table 4. Number of Phaseolus vulgaris plants (two varieties pooled) showing damage (0-5 rating) when infested at two 19

phenological stages (12 and 13, first and third true leaf, respectively) with either 5 or 10 Delia platura larvae in a growth 20

chamber experiment, n≈12 for each treatment. Significant difference observed between first and third true leaf phenological

Comment [BS11]: Comment 15

22 stages (*P* < 0.05, Residual Deviance 186.3, AIC 204.3).

21

Plant Growth				Damage rating	e rating	50		
Stage	Larvae	0	-	2	m	4	S	
12	0	12	0	0	0	0	0	
(First true leaf)	5	1	0	-	3	2	3	
	10	0	0	1	7	7	7	
13	0	12	0	0	0	0	0	
Third true leaf)	5	5	9	0	0	0	-	
	10	2	2	3	1	4	0	

23

Cambridge University Press

- 1 Figure captions
- 2 Figure 1. Plot design for 2016 field trials at Canning, Nova Scotia and
- 3 Wilmot Valley, PEI. Letters A through D represent planting dates. Dark grey
- 4 areas indicate guard plants (Gold Rush P. vulgaris var). Numbered boxes
- 5 show location of blue sticky traps for monitoring adult populations of Delia
- 6 spp.
- 7 Figure 2. Mean number of D. platura and D. florilega females (A) and D.
- 8 platura males (B) and D. florilega males (C), identified per trap from four
- 9 traps collected weekly from organic commercial bean fields on Prince
- 10 Edward Island (2015 and 2016) and Nova Scotia (2016).

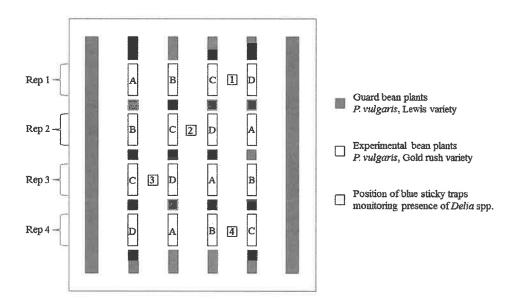
11

- Figure 3: Mean number (± SE) D. platura eggs per plant when given choice
- of two varieties of P. vulgaris: Gold Rush and Lewis varieties in a growth
- 14 chamber study. Phenological growth stage (varieties pooled) followed by the
- same letter not significantly different (P < 0.05). For most growth stages
- n=10, with the exception of growth stages 12 and 13 (n = 9) and 75 (n = 8).

17

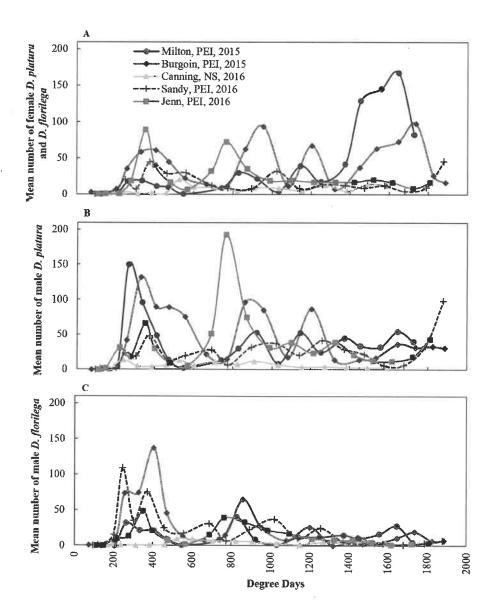
- 18 Figure 4: Mean number of D. platura and D. florilega females per blue
- 19 sticky trap from organic bean fields in Canning, Nova Scotia (A), Wilmot
- 20 Valley (B) and Saint Ann (C) on PEI in 2016 by accumulated degree days

- 21 (base 3.9, start date of 1 January). Numbers 1-5 denote planting times in
- 22 conjunction with Delia phenology.



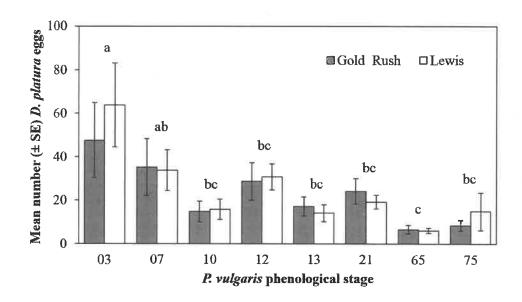
Plot design for 2016 field trials at Canning, Nova Scotia and Wilmot Valley, PEI. Letters A through D represent planting dates. Dark grey areas indicate guard plants (Gold Rush P. vulgaris var). Numbered boxes show location of blue sticky traps for monitoring adult populations of Delia spp.

158x94mm (240 x 240 DPI)



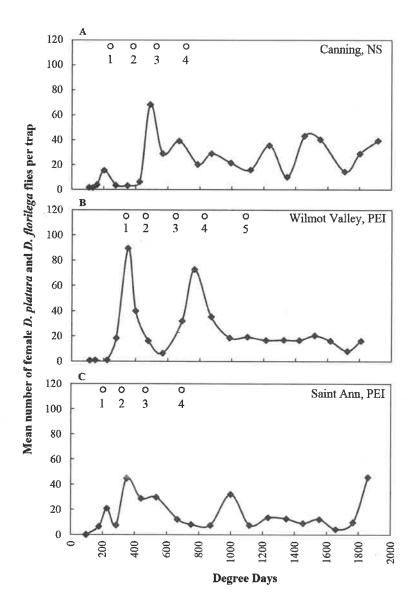
Mean number of D. platura and D. florilega females (A) and D. platura males (B) and D. florilega males (C), identified per trap from four traps collected weekly from organic commercial bean fields on Prince Edward Island (2015 and 2016) and Nova Scotia (2016).

169x207mm (240 x 240 DPI)



Mean number (\pm SE) D. platura eggs per plant when given choice of two varieties of P. vulgaris: Gold Rush and Lewis varieties in a growth chamber study. Phenological growth stage (varieties pooled) followed by the same letter not significantly different (P < 0.05). For most growth stages n=10, with the exception of growth stages 12 and 13 (n = 9) and 75 (n = 8).

147x87mm (240 x 240 DPI)



Mean number of D. platura and D. florilega females per blue sticky trap from organic bean fields in Canning, Nova Scotia (A), Wilmot Valley (B) and Saint Ann (C) on PEI in 2016 by accumulated degree days (base 3.9, start date of 1 January). Numbers 1-5 denote planting times in conjunction with Delia phenology.

136x202mm (240 x 240 DPI)